Nair P, Gaga M, Zervas E, et al. Safety and efficacy of a CXCR2 antagonist in patients with severe asthma and sputum neutrophils: a randomized, placebo-controlled clinical trial. Clin Exp Allergy 2012;42:1097-103.
PDGF rece trol in se
2009;64:11
12. Busse

4. Wenzel S, Ford L, Pearlman D, et al. Dupilumab in persistent asthma with elevated eosinophil levels. N Engl J Med 2013;368: 2455-66.

5. Pavord ID, Korn S, Howarth P, et al. Mepolizumab for severe eosinophilic asthma (DREAM): a multicenter, double-blind, placebo-controlled trial. Lancet 2012;380:651-9.

6. Nair P, Pizzichini MM, Kjarsgaard M, et al. Mepolizumab for prednisone-dependent asthma with sputum eosinophilia. N Engl J Med 2009;360:985-93.

 Haldar P, Brightling CE, Hargadon B, et al. Mepolizumab and exacerbations of refractory eosinophilic asthma. N Engl J Med 2009;360:973-84. [Erratum, N Engl J Med 2011;364:588.]
Castro M, Mathur S, Hargreave F, et al. Reslizumab for poorly controlled, eosinophilic asthma: a randomized, placebocontrolled study. Am J Respir Crit Care Med 2011;184:1125-32.
Corren J, Lemanske RF, Hanania NA, et al. Lebrikizumab treatment in adults with asthma. N Engl J Med 2011;365:1088-98.
Piper E, Brightling C, Niven R, et al. A phase II placebocontrolled study of tralokinumab in moderate-to-severe asthma. Eur Respir J 2013;41:330-8.

11. Humbert M, de Blay F, Garcia G, et al. Masitinib, a c-kit/

PDGF receptor tyrosine kinase inhibitor, improves disease control in severe corticosteroid-dependent asthmatics. Allergy 2009;64:1194-201.

12. Busse WW, O'Byrne PM, Bleecker ER, et al. Safety and tolerability of the novel inhaled corticosteroid fluticasone furoate in combination with the β 2 agonist vilanterol administered once daily for 52 weeks in patients \geq 12 years old with asthma: a randomised trial. Thorax 2013 February 25 (Epub ahead of print).

13. Busse WW, Wenzel SE, Meltzer EO, et al. Safety and efficacy of the prostaglandin D2 receptor antagonist AMG 853 in asthmatic patients. J Allergy Clin Immunol 2013;131:339-45.

14. Barnes N, Pavord I, Chuchalin A, et al. A randomized, double-blind, placebo-controlled study of the CRTH2 antagonist OC000459 in moderate persistent asthma. Clin Exp Allergy 2012;42:38-48.

15. Beeh KM, Kanniess F, Wagner F, et al. The novel TLR-9 agonist QbG10 shows clinical efficacy in persistent allergic asthma. J Allergy Clin Immunol 2013;131:866-74.

16. Wechsler ME, Fulkerson PC, Bochner BS, et al. Novel targeted therapies for eosinophilic disorders. J Allergy Clin Immunol 2012;130:563-71.

DOI: 10.1056/NEJMe1305426 Copyright © 2013 Massachusetts Medical Society

Releasing the Brake on Puberty

Ieuan A. Hughes, M.D.

What is so magical about the age at onset of puberty in humans — currently set at approximately 11 years of age?1 Why not 6 or 16? Indeed, addressing this question from the perspective of evolutionary biology² suggests that puberty, as defined by age at menarche in girls, was earlier in Neolithic times and became delayed during the Industrial Revolution before reverting over the past two centuries to the present set point.³ The shortened life span and the need to reach reproductive capacity would seem to have been predominant influences on the age of menarche thousands of years ago, whereas the more recent fluctuations in the age at onset of puberty have been attributed to poor nutrition followed by social improvement. How can one explain the underlying cause of the early onset of puberty in a child brought to the clinic at 5 years of age? It clearly cannot be explained on environmental grounds, even allowing for the secular trends in puberty ascribed to increasing obesity in the childhood population.4,5 Influences on the timing of puberty, for the most part, remain unknown; the endocrinologist still cannot explain simply to parents why puberty generally starts at the age of 11 years, let alone why their child has entered puberty at 6 years of age.

The role of genetic factors in the control of the onset of puberty is vividly illustrated by a report in this issue of the Journal of a familial form of precocious puberty caused by loss-offunction mutations in an imprinted gene.6 The authors had access to 40 members of 15 families in whom affected probands had central precocious puberty - that is, premature reactivation of the pulse GnRH generator that underscores the onset of normal puberty.7 Applying wholeexome sequencing in multiply affected families adequately phenotyped for central precocious puberty, the authors identified deleterious mutations in a paternally expressed imprinted gene, MKRN3. (Imprinted genes have a "sex bias" in that they are expressed only from the maternal or the paternal chromosome; some genes are paternally imprinted, whereas others are maternally imprinted. MKRN3 is maternally imprinted; expression from the maternally inherited copy of the gene is suppressed. MKRN3 protein is thus derived from RNA transcribed exclusively from the paternally inherited copy of the gene.)

MKRN3 encodes makorin RING-finger protein 3, which is involved with ubiquitination and cell signaling. The makorin family of proteins is abundantly expressed in the developing brain,

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on July 16, 2013. For personal use only. No other uses without permission.

Copyright © 2013 Massachusetts Medical Society. All rights reserved.



Figure 1. Timing of Puberty.

A pivotal event in the onset of puberty in mammals is the resumption of pulsatile release of gonadotropin-releasing hormone (GnRH) from neurons of the hypothalamus. Known influences on the timing of the onset of puberty in mammals include the photoperiod, leptin levels, and the increased expression of neurokinin B, kisspeptin, and their receptors (NK3R and KISS1R, respectively). Abreu et al.⁶ implicate MKRN3, a protein that is believed to mediate ubiquitination, in puberty onset. In contrast with kisspeptin and neurokinin B, which stimulate the commencement of puberty, MKRN3 seems to inhibit puberty: Abreu et al. show that mutations in *MKRN3* predicted to cause loss of function of the protein cause central precocious puberty. KNDy denotes kisspeptin–neurokinin B–dynorphin, INF infundibular nucleus, ME median eminence, and POA preoptic area.

including the arcuate nucleus, where there is a repository of genes whose expression is relevant to puberty.^{8,9} The authors showed that the expression of *Mkrn3* in mice of both sexes was highest at postnatal day 10 and declined thereafter to reach a nadir precisely consonant with

the onset of puberty. It is also at this point that the expression of genes considered central to the activation of puberty (e.g., the kisspeptins and neurokinin B) begins to increase.¹⁰

So what is the effect of this study of familial central precocious puberty on our knowledge about the way in which the onset of puberty is controlled in humans? More is known about why puberty may be delayed than why it commences precociously. Any chronic disease process, such as severe malnutrition or a systemic disease such as cystic fibrosis, will delay or halt the progression of puberty. More specifically, the hypothalamic-pituitary-gonadal endocrine pathway can harbor specific defects in hormone production that are the result of known mutations. Hypogonadotropic hypogonadism caused by loss-of-function mutations affecting the G-protein-coupled receptor KISS1R (also known as GPR54) and those involving neurokinin B result in failure of the normal pattern of pulsatile GnRH secretion required to stimulate gonadotropin production and subsequently gonadal steroid secretion.^{11,12} The discovery of the effect of mutations in MKRN3 in humans and of a suggested role for its mouse orthologue in the arcuate nucleus appears to cement the idea that puberty starts only with the release of a restraint mechanism on the GnRH pulse generator, which in turn releases the brake on puberty (Fig. 1). A release of this restraint mechanism probably also explains why intracranial damage from conditions as diverse as head trauma and hydrocephalus and the effects of cranial irradiation can lead to precocious puberty.

Although the finding of a genetic cause for central precocious puberty is a significant contribution to further understanding human puberty, an explanation is lacking about why puberty starts at about the time of the junction of the first and second decades of human life. How MKRN3, an exemplar of a neurobiologic brake, interacts with other major players of puberty, such as kisspeptin, GnRH, leptin, and a host of neurotransmitters (excitatory and inhibitory), will certainly continue to exercise the minds of the puberty pundits.

Disclosure forms provided by the author are available with the full text of this article at NEJM.org.

From the University of Cambridge, Cambridge, United Kingdom.

This article was published on June 5, 2013, at NEJM.org.

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on July 16, 2013. For personal use only. No other uses without permission.

Copyright © 2013 Massachusetts Medical Society. All rights reserved.

 Styne DM, Grumbach MM. Puberty: ontogeny, neuroendocrinology, physiology and disorders. In: Melmed S, Polonsky KS, Larsen PR, Kronenberg HM, eds. Williams textbook of endocrinology. 12th ed. Philadelphia: Elsevier Saunders, 2001:1054-201.
Gluckman PD, Bergstrom CT. Evolutionary biology within medicine: a perspective of growing value. BMJ 2011;343:d7671.
Gluckman PD, Hanson MA. Evolution, development and timing of puberty. Trends Endocrinol Metab 2006;17:7-12.

4. Marcovecchio ML, Chiarelli F. Obesity and growth during childhood and puberty. World Rev Nutr Diet 2013;106:135-41.

5. Cousminer DL, Berry DJ, Timpson NJ et al. Genome-wide association and longitudinal analyses reveal genetic loci linking height growth, pubertal timing and childhood adiposity. Hum Mol Genet 2013 March 21 (Epub ahead of print).

6. Abreu AP, Dauber A, Macedo DB, et al. Central precocious puberty caused by mutations in the imprinted gene *MKRN3*. N Engl J Med 2013;368:2467-75.

7. Knobil E. The GnRH pulse generator. Am J Obstet Gynecol 1990;163:1721-7.

8. Gray TA, Hernandez L, Carey AH, et al. The ancient source of a distinct gene family encoding proteins featuring RING and C(3)H zinc-finger motifs with abundant expression in developing brain and nervous system. Genomics 2000;66:76-86.

9. Goodman RL, Lehman MN. Kisspeptin neurons from mice to men: similarities and differences. Endocrinology 2012; 153:5105-18.

10. George JT, Seminara SB. Kisspeptin and the hypothalamic control of reproduction: lessons from the human. Endocrinology 2012;153:5130-6.

11. Topaloglu AK, Tello JA, Kotan LD, et al. Inactivating KISS1 mutation and hypogonadotropic hypogonadism. N Engl J Med 2012;366:629-35.

12. Topaloglu AK, Reimann F, Guclu M, et al. TAC3 and TACR3 mutations in familial hypogonadotropic hypogonadism reveal a key role for neurokinin B in the central control of reproduction. Nat Genet 2009;41:354-8.

DOI: 10.1056/NEJMe1306743 Copyright © 2013 Massachusetts Medical Society.

G Proteins — The Disease Spectrum Expands

Allen M. Spiegel, M.D.

are heterotrimers composed of guanosine triphosphate-binding alpha subunits and tightly linked beta and gamma subunits. They couple a vast array of receptors (G-protein-coupled receptors, the subject of the Nobel Prize in Chemistry this past year¹) to effectors that regulate diverse cellular processes. Of 15 human alpha-subunit genes, some, such as GNAS, are expressed ubiquitously; others are expressed only in specialized cells. G_s (the G protein encoded by GNAS) couples many hormone and neurotransmitter receptors to cyclic AMP stimulation, and it was the first G protein to be associated with human disease. Germline mutations that inactivate $G\alpha_s$ (the G-protein subunit α_s) were shown to cause the prototypical hormone-resistance disorder, pseudohypoparathyroidism.² Somatic activating mutations cause sporadic endocrine tumors and the McCune-Albright syndrome.² Mutations subsequently were identified in genes that encode dysfunctional G α proteins in rod photoreceptors in forms of night blindness and in cone photoreceptors in forms of color blindness, and mutations in GNAL have been linked to primary torsion dystonia.3

Somatic mutations that activate $G\alpha_q$ (the G-protein subunit α_q) and $G\alpha_{11}$ (the G-protein subunit α_{11}), closely related G proteins that activate intracellular ionized calcium–mediated signaling, have been associated with uveal melanoma.⁴ In this issue of the *Journal*, Nesbit⁵ and Mannstadt⁶ and their colleagues report that germline muta-

G proteins (guanine nucleotide-binding proteins) tions that inactivate $G\alpha_{11}$ cause hypercalcemic are heterotrimers composed of guanosine triphosphate-binding alpha subunits and tightly $G\alpha_{11}$ cause hypocalcemic disorders.

> Calcium homeostasis is tightly regulated by parathyroid hormone. Parathyroid hormone secretion from the parathyroid glands is inhibited directly by increased serum levels of calcium. Primary hyperparathyroidism, the major cause of hypercalcemia in patients seen in an ambulatory setting, is caused by a neoplastic process in one or more parathyroid glands. This process leads to excess parathyroid hormone secretion, despite increased serum levels of calcium.

> Familial hypocalciuric hypercalcemia is an autosomal dominant disease that, like primary hyperparathyroidism, is characterized by hypercalcemia and normal or elevated levels of serum parathyroid hormone.⁷ However, renal and skeletal manifestations of primary hyperparathyroidism are generally absent in patients with familial hypocalciuric hypercalcemia. Partial parathyroidectomy does not correct the hypercalcemia; thus, surgery is not indicated. The patient's family history and measurement of urinary calcium:creatinine ratios are key to distinguishing familial hypocalciuric hypercalcemia from primary hyperparathyroidism.

> Previous studies have shown that many cases of familial hypocalciuric hypercalcemia are caused by heterozygous germline inactivating mutations in the gene encoding the calcium-sensing receptor (*CASR*). The calcium-sensing receptor is a G-protein-coupled receptor that is highly ex-

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on July 16, 2013. For personal use only. No other uses without permission.

Copyright © 2013 Massachusetts Medical Society. All rights reserved.