REVIEW ARTICLE

MEDICAL PROGRESS

Julie R. Ingelfinger, M.D., Editor

Primary Hyperoxaluria

Pierre Cochat, M.D., Ph.D., and Gill Rumsby, Ph.D.

THE PRIMARY HYPEROXALURIAS ARE a group of autosomal recessive disorders involving the overproduction of oxalate. Although the initial recognition of the disease is attributed to Lepoutre, who reported it in 1925,¹ the elucidation of the underlying biochemical abnormalities occurred many years later. This review discusses the major biochemical, genetic, and therapeutic advances that have led to a better understanding of primary hyperoxaluria.

Oxalate, a dicarboxylic acid (HOOC-COOH), is a highly insoluble end product of metabolism in humans. It is excreted almost entirely by the kidney, particularly in the form of its calcium salt, and has a tendency to crystallize in the renal tubules. The main defect of inherited hyperoxaluria is the overproduction of oxalate, primarily by the liver, which results in increased excretion by the kidney. The earliest symptoms among those affected are urolithiasis and nephrocalcinosis, which lead to progressive renal involvement and chronic kidney disease. Renal damage is ultimately caused by a combination of tubular toxicity from oxalate, nephrocalcinosis (with both intratubular and interstitial deposits of calcium oxalate), and renal obstruction by stones, often with superimposed infection. Inflammation has recently been shown to contribute to the progression of chronic kidney disease in animal models of nephrocalcinosis induced by calcium oxalate.² A second phase of damage that is the result of primary hyperoxaluria occurs when the glomerular filtration rate (GFR) drops to 30 to 45 ml per minute per 1.73 m² of body-surface area and the kidney is unable to effectively excrete the oxalate load it receives. At this point, plasma levels of oxalate rise and exceed saturation,³ and oxalate is subsequently deposited in all tissues (systemic oxalosis), particularly in the skeleton.

Secondary hyperoxaluria may occur as a result of excess dietary intake or poisoning with oxalate precursors or may be the result of enteric hyperoxaluria. The latter can occur after bowel resection, which can lead to sequestration of calcium in the gut, leaving oxalate in its more soluble sodium form, which is then taken up by the colon. Secondary hyperoxaluria must be ruled out before an investigation for primary hyperoxaluria begins.

EPIDEMIOLOGY

The true prevalence of primary hyperoxaluria is unknown. Primary hyperoxaluria type 1, the most common form, has an estimated prevalence of 1 to 3 cases per 1 million population and an incidence rate of approximately 1 case per 120,000 live births per year in Europe.^{4,5} It accounts for 1 to 2% of cases of pediatric end-stage renal disease (ESRD), according to registries from Europe, the United States, and Japan,⁶ but it appears to be more prevalent in countries in which consanguineous marriages are common (with a prevalence of 10% or higher in some North African and Middle Eastern nations).⁷

From Centre de Référence des Maladies Rénales Rares Néphrogones; Centre de la Recherche Scientifique — Unité Mixte de la Recherche 5305, Hospices Civils de Lyon, and Université Claude-Bernard Lyon 1 — all in Lyon, France (P.C.); and Clinical Biochemistry, University College London Hospitals, London (G.R.). Address reprint requests to Dr. Cochat at Service de Néphrologie Rhumatologie Dermatologie Pédiatriques, Hôpital Femme Mère Enfants, 59 Blvd. Pinel, 69677 Bron CEDEX, France, or at pierre.cochat@chu-lyon.fr.

N Engl J Med 2013;369:649-58. DOI: 10.1056/NEJMra1301564 Copyright © 2013 Massachusetts Medical Society.

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on September 13, 2013. For personal use only. No other uses without permission.

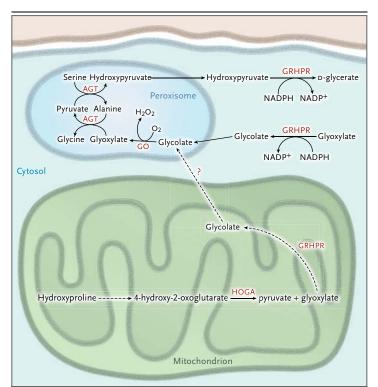


Figure 1. Glyoxylate Metabolism in the Hepatocyte.

In the normal hepatocyte, peroxisomal alanine-glyoxylate aminotransferase (AGT) catalyzes the conversion of glyoxylate and alanine to pyruvate and glycine and of serine to hydroxypyruvate. Glyoxylate can also be used by cytosolic glyoxylate reductase–hydroxypyruvate reductase (GRHPR) to produce glycolate, and this enzyme also catalyzes the metabolism of hydroxypyruvate to D-glycerate, in both cases with NADPH as cofactor. Mitochondrial glyoxylate is produced when collagen is broken down to hydroxyproline and 4-hydroxy-2-oxoglutarate aldolase (HOGA), which catalyzes the last step in this pathway. The fate of mitochondrial glyoxylate has not been completely delineated; it may be metabolized by mitochondrial GRHPR. In primary hyperoxaluria types 1 and 2, glyoxylate accumulates as a result of AGT and GRHPR deficiency, respectively, and is converted to oxalate by lactate dehydrogenase. Primary hyperoxaluria type 3 results from a defect in HOGA, although it is not clear why this defect would cause an increase in oxalate levels. Glycolate oxidase (GO) is a potential target for substrate reduction therapy.

FORMS OF PRIMARY HYPEROXALURIA

There are three forms of primary hyperoxaluria in which the underlying defects have been identified; they are designated as primary hyperoxaluria types 1, 2, and 3. Each is caused by an enzyme deficiency, and each affects a different intracellular organelle (Fig. 1). Common to all is the overproduction of oxalate. In the case of primary hyperoxalurias 1 and 2, the mechanism is the action of lactate dehydrogenase on the immediate oxalate precursor, glyoxylate. The source of oxalate in primary hyperoxaluria 3 has not been identified.

Primary hyperoxaluria type 1 (number 259900 in the Online Mendelian Inheritance of Man [OMIM] database) is caused by a deficiency of the liver-specific peroxisomal enzyme alanineglyoxylate aminotransferase (AGT), a pyridoxal 5'-phosphate-dependent enzyme that catalyzes the transamination of glyoxylate to glycine. This deficiency results in the accumulation of glyoxylate and excessive production of both oxalate and glycolate. AGT is a stable homodimer, with its N-terminal amino acids wrapped around the adjacent monomer.⁸ A common variant, Pro-11Leu, creates a stronger N-terminal mitochondrial targeting sequence, which influences the fate of some mutant proteins.

Primary hyperoxaluria type 2 (OMIM number, 260000) is caused by a lack of glyoxylate reductase–hydroxypyruvate reductase (GRHPR), which catalyzes the reduction of glyoxylate to glycolate and hydroxypyruvate to D-glycerate. GRHPR has a wide tissue distribution, but it is primarily intrahepatic,⁹ present largely in the cytosol of hepatocytes and to a lesser extent in mitochondria.^{10,11} When there is a deficiency of GRHPR, lactate dehydrogenase metabolizes the accumulated glyoxylate to oxalate and the hydroxypyruvate to L-glycerate.

Primary hyperoxaluria type 3 (OMIM number, 613616) results from defects in the liverspecific mitochondrial enzyme 4-hydroxy-2-oxoglutarate aldolase (HOGA).¹² This enzyme plays a key role in the metabolism of hydroxyproline, and kinetic studies suggest that the forward reaction, in which 4-hydroxy-2-oxoglutarate (HOG) is converted to pyruvate and glyoxylate, is favored.¹³ However, it is unclear why this defect would cause an increase in oxalate levels, since impairment of the synthesis of mitochondrial glyoxylate would be expected. One theory is that the substrate HOG breaks down to oxalate either enzymatically¹⁴ or in some other way; another is that HOG inhibits mitochondrial GRHPR.¹⁵

GENETIC FEATURES

Mutations in *AGXT*, the gene encoding AGT, result in primary hyperoxaluria type 1. One mutation, Gly170Arg, can lead to significant catalytic

N ENGLJ MED 369;7 NEJM.ORG AUGUST 15, 2013

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on September 13, 2013. For personal use only. No other uses without permission.

activity in vitro, but in some cases remains at the low end of the normal reference range.16 This mutation and three others (Ile244Thr, Phe152Ile, and Gly41Arg) unmask the N-terminal mitochondrial targeting sequence of AGT (encoded by Pro11Leu), leading to peroxisome-to-mitochondrion mistargeting (in which AGT, which normally targets peroxisomes, instead targets mitochondria).17 At least 178 mutations have been described18; of these, Gly170Arg and c.33dupC occur across populations at a frequency of 30% and 11%, respectively.¹⁹ In contrast, the Ile244Thr mutation is especially common in North Africa and Spain.^{20,21} The only known aspect of primary hyperoxaluria type 1 with a strong genotype-phenotype relationship is responsiveness to pyridoxine, which occurs in patients with the Gly170Arg and Phe152Ile mutations^{22,23} and is mediated by effects on protein stability, catalytic activity, and peroxisomal import.24 There is no association between age at onset and mutation, and there can be marked intrafamilial clinical heterogeneity.25

A total of 30 mutations have been identified in *GRHPR*, the gene that is defective in primary hyperoxaluria type 2,¹⁸ and 2 of these mutations, c.103delG and c.403_404+2delAAGT, are relatively common; the former occurs exclusively in white populations, the latter almost entirely in Asian populations.⁹

The association of mutations in *HOGA1* with primary hyperoxaluria type 3 has been reported recently,¹² and 19 mutations have been described.^{12,26-28} One particular variant, c.700+5G \rightarrow T, accounts for half of all mutant alleles.

CLINICAL SPECTRUM

Primary hyperoxaluria may occur at almost any age — from birth to the sixth decade of life — with a median age at onset of 5.5 years.²⁹ The clinical presentation varies from infantile nephrocalcinosis and failure to thrive as a result of renal impairment to recurrent or only occasional stone formation in adulthood. However, 20 to 50% of patients have advanced chronic kidney disease or even ESRD at the time of diagnosis.^{6,29,30} Roughly 10% of patients receive a diagnosis of primary hyperoxaluria only when the disease recurs after kidney transplantation. In other cases, the disease is identified before symptoms appear in the course of family evalua-

tions. Kidney injury, leading to a decrease in the GFR, results in chronic kidney failure and ultimately in ESRD, together with progressive systemic involvement (Fig. 2). The major sites of crystal deposition are the kidneys, the bloodvessel walls, and the bones, with crystal deposits often leading to fractures. Oxalosis can also affect the joints, retina,³¹ skin, bone marrow,³² heart,33 and central nervous system,34 leading to severe illness and death. Data from the Rare Kidney Stone Consortium indicate that the median age at diagnosis of ESRD is 24 years.³⁵ According to the European pediatric registry, the median age at the initiation of renal-replacement therapy is 1.5 years, and the patient survival rate 5 years after the initiation of renal-replacement therapy is 76%, as compared with 92% among children with ESRD resulting from other conditions. These figures translate into a risk of death for patients with primary hyperoxaluria that is three times as high as the risk for those without the disease.6

Primary hyperoxaluria type 1 is the most devastating subtype, particularly when it occurs in infancy, but patients who have the Gly170Arg or Phe152Ile mutation have a better overall outcome than other patients with type 1 disease, partly because of their sensitivity to pyridoxine.22,36 Patients with primary hyperoxaluria type 2 appear to have a less severe course, although the two disorders cannot be distinguished according to age at onset, and in some instances, primary hyperoxaluria type 2 is initially assumed to be type 1.37 Primary hyperoxaluria type 3 has the least severe course and may be silent or limited to stone formation, sometimes even improving over time.27 Whereas hyperoxaluria persists in primary hyperoxaluria type 3, nephrocalcinosis and chronic kidney failure are uncommon, and systemic involvement has not been reported thus far. Other factors, including environmental factors and modifier genes, may contribute to the clinical heterogeneity of primary hyperoxaluria.

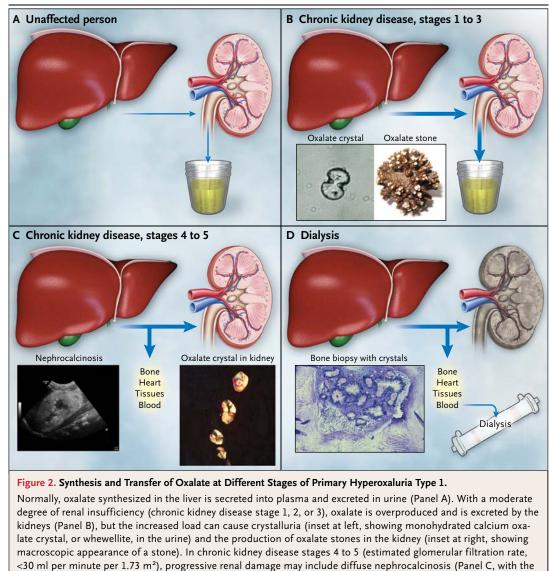
DIAGNOSIS

Given its rarity, primary hyperoxaluria may go unrecognized for several years after the onset of symptoms. Considering the possibility of primary hyperoxaluria and pursuing an evaluation that

651

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on September 13, 2013. For personal use only. No other uses without permission.



inset at left showing diffuse nephrocalcinosis on an ultrasonogram in a 3-month-old infant and the inset at right showing oxalate crystals in the proximal renal tubule under polarized light). In patients receiving dialysis, the oxalate load cannot be cleared effectively, and oxalate crystals are deposited in bones, the heart, and other tissues (Panel D, with inset showing oxalate crystals in a bone-biopsy specimer; May–Grünwald–Giemsa stain). (Panel C insets are courtesy of Dr. Frederique Dijoud, and Panel D inset is courtesy of Dr. Georges Boivin.)

is in accordance with published algorithms may facilitate earlier recognition.^{38,39}

Because a majority of patients with primary hyperoxaluria present with symptoms related to urolithiasis, assessment of the risk of kidney stones, based on measurements of urinary levels of oxalate, calcium, citrate, sodium, magnesium, and urate, as well as urinary pH and volume, is central to a good evaluation. In patients with primary hyperoxaluria, kidney stones usually

consist of more than 95% calcium oxalate monohydrate (whewellite), and they are unusually pale in color and nonhomogeneous in appearance.^{40,41} A finding of oxalate crystals in a kidney-biopsy specimen is also suggestive of primary hyperoxaluria (Fig. 2). In infancy, the chief presenting feature is metabolic acidosis, along with acute renal failure.

central to a good evaluation. In patients with The excretion of urinary oxalate is variable, primary hyperoxaluria, kidney stones usually particularly in the first year of life (Table 1), but

N ENGLJ MED 369;7 NEJM.ORG AUGUST 15, 2013

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on September 13, 2013. For personal use only. No other uses without permission.

persistently elevated excretion (>0.7 mmol per 1.73 m² per day,³⁸ or a urinary oxalate:creatinine ratio greater than the reference range for age), in addition to suggestive clinical symptoms and the absence of secondary hyperoxaluria, indicates the need for further evaluation. Not all patients with primary hyperoxaluria have markedly elevated levels of urinary oxalate, but if symptoms are suggestive, an additional evaluation for primary hyperoxaluria should be considered. Measurements of other urinary metabolites, such as glycolate and L-glycerate, are helpful but nonspecific. Levels of glycolate are elevated in two thirds of patients with primary hyperoxaluria type 1 and may also be elevated in patients with type 3.26 An increased L-glycerate level was formerly regarded as pathognomonic for primary hyperoxaluria type 2,45 but the level is not invariably elevated.46 The presence of precursors of HOG has recently been reported in the urine of patients with primary hyperoxaluria type 3,14,15 and the development of routine methods for the detection of such precursors may facilitate the diagnosis of primary hyperoxaluria in a wider group of patients who have kidney stones. Whereas urinary calcium levels are typically low in primary hyperoxaluria types 1 and 2, levels in type 3 may vary and in some instances are quite high.^{26,27,42} Measurement of plasma levels of oxalate should be reserved for patients with stage 3b chronic kidney disease (estimated GFR, 30 to 45 ml per minute per 1.73 m²), since plasma levels remain relatively normal until kidney function is substantially impaired. Attempts have been made to establish a threshold plasma oxalate level that can be used to differentiate between primary hyperoxaluria and kidney failure from any cause.3,43 There is considerable overlap in plasma oxalate values among kidney diseases, although in our opinion, values greater than 50 μ mol per liter are suggestive of primary hyperoxaluria.

A definitive diagnosis of primary hyperoxaluria in a patient with clinical signs and symptoms requires genetic testing. In the absence of any additional information, it seems reasonable to conduct testing for type 1 first, since it accounts for approximately 80% of cases of primary hyperoxaluria.⁴² If there is no genetic evidence of type 1 disease, the next step is to look for mutations indicating type 2 or type 3, which have a similar frequency. The strategy for ge-

Table 1. Age-Related Reference Ranges of Metabolites in Patients with Primary Hyperoxaluria.*			
Urinary Excretion	Reference Range	Source	
24-Hr specimen			
Oxalate, all ages	<45 mg (0.5 mmol)/1.7 m²	Hoppe ⁴²	
Glycolate, all ages	<45 mg (0.5 mmol)/1.73 m²	Hoppe ⁴²	
Random ("spot") specimen			
Oxalate:creatinine		Barratt et al. ⁴³	
<l td="" yr<=""><td>11.9–207 μg/mg (15–260 μmol/mmol)</td><td></td></l>	11.9–207 μg/mg (15–260 μmol/mmol)		
1 to <5 yr	8.7–95.6 μg/mg (11–120 μmol/mmol)		
5 to 12 yr	47–119 μg/mg (60–150 μmol/mmol)		
>12 yr	1.6–63.7 μg/mg (2–80 μmol/mmol)		
Glycolate:creatinine		Barratt et al.43	
<l td="" yr<=""><td>5.4–47.0 μg/mg (8–70 μmol/mmol)</td><td></td></l>	5.4–47.0 μg/mg (8–70 μmol/mmol)		
1 to <5 yr	4.0–61.4 μg/mg (6–91 μmol/mmol)		
5 to 12 yr	4–31 μg/mg (6–46 μmol/mmol)		
>12 yr	2.7–27.0 μg/mg (4–40 μmol/mmol)		
Glycerate:creatinine		Dietzen et al. ⁴⁴	
0 to 5 yr	12–177 μg/mg (13–190 μmol/mmol)		
>5 yr	19–115 μg/mg (22–123 μmol/mmol)		
HOG:creatinine, adults	0.1–3.9 μg/mg (0.07–2.8 μmol/mmol)	Belostotsky et al. ¹⁴	

* Actual data are assay-dependent; these data are intended to provide a guide for clinicians. HOG denotes 4-hydroxy-2-oxoglutarate.

netic testing should also take race and ethnic group into account. The disadvantage of limiting testing to genetic studies is the absence of functional analysis (not all genetic variants are pathologic). In addition, deletions may not be detected unless parental studies are also performed. In some cases, the phenotype is typical of primary hyperoxaluria, but no mutation is detected, either because the mutation lies in a promoter or other regulatory sequence or because some other, as yet undefined, metabolic defect is present (i.e., "uncategorized" primary hyperoxaluria). In such cases, a liver biopsy can

653

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on September 13, 2013. For personal use only. No other uses without permission.

be performed to test for levels of AGT and GRHPR activity; if the results are negative, primary hyperoxaluria types 1 and 2 can be ruled out. As yet there is no enzymatic test that can be used to diagnose type 3, although tests showing elevated hepatic levels of HOGA have been described.¹⁵

For persons with a family history of primary hyperoxaluria, particularly type 1, genetic screening can be performed, and testing during the first trimester of pregnancy can establish a prenatal diagnosis.⁴⁷ Preimplantation diagnosis may be possible, depending on local facilities.

MANAGEMENT

SUPPORTIVE MEASURES

Once a diagnosis of primary hyperoxaluria is being considered, supportive measures should be initiated, since long-term adherence to such treatment can dramatically improve the prognosis and slow the progression to ESRD.48 Fluid intake of more than 2 to 3 liters per square meter of body-surface area per day is essential for stone prevention,³⁹ but in infants tube or gastrostomy feeding may be required to obtain appropriately dilute urine around the clock. Oral potassium citrate (0.10 to 0.15 mg per kilogram of body weight per day) is used to alkalinize urine (ideal pH, 6.2 to 6.8) and, more important, to inhibit crystallization⁴²; if renal function is impaired, sodium citrate should be used to avoid an increase in the potassium load.49

Pyridoxine supplementation is helpful in primary hyperoxaluria type 1 (but not in other forms of primary hyperoxaluria). A starting dose of 5 mg per kilogram per day may be progressively increased but should not exceed 20 mg per kilogram, with a trial period of at least 3 months.39 Responsiveness is defined as a decrease in the level of urinary oxalate by more than 30% from the point of treatment initiation. As previously mentioned, the Gly170Arg and Phe152Ile genotypes are associated with a significant and sustained reduction of urinary oxalate levels during treatment with pyridoxine, which leads to improvement in the overall prognosis.23 Responsiveness to pyridoxine has also been observed in patients with the Ile244Thr genotype.³⁰ Pyridoxine can be discontinued after 3 months if oxalate levels have not fallen, provided that complete adherence to treatment has been confirmed.

The intestinal oxalate load has a limited effect on disease progression in primary hyperoxaluria, since the main source of oxalate is endogenous. Consequently, oxalate-rich foods should be restricted only as a precaution,⁵⁰ and normal calcium intake should be maintained. Probiotics that break down oxalate (e.g., *Oxalobacter formigenes*) may have a role in promoting intestinal oxalate excretion,⁵¹ although a recent clinical trial had disappointing results.⁵²

Extracorporeal shock-wave lithotripsy (ESWL) is not recommended in patients with primary hyperoxaluria who have a heavy stone burden, both because calcium oxalate stones do not easily fragment^{53,54} and because the risk of parenchymal damage, particularly in small kidneys, is high (a problem that has been reported in studies of primary hyperoxaluria in animals).⁵⁵ For affected patients with a high stone burden, minimally invasive methods (e.g., ureteroscopic laser lithotripsy with percutaneous stone removal) are preferable to ESWL.^{53,56}

DIALYSIS

Conventional hemodialysis and peritoneal dialysis do not eliminate sufficient levels of oxalate to avert a continuous positive balance (Fig. 2D). Thus, more intensive strategies must be used to clear plasma oxalate levels and to limit systemic involvement.⁵⁷ If preemptive transplantation is not feasible, therapeutic strategies that include short daily sessions of high-flux dialysis, nocturnal dialysis, or combinations of hemodialysis and nocturnal peritoneal dialysis are needed to keep predialysis levels of plasma oxalate below 30 to 45 μ mol per liter.^{34,58}

If a patient who presents with ESRD is found to have primary hyperoxaluria type 1, responsiveness to pyridoxine should be tested, since a pyridoxine-induced reduction in plasma oxalate levels would influence the dialysis strategy.³⁴ Once kidney failure occurs, oxalate deposition in the bone marrow may result in treatment-resistant anemia.⁵⁹ In addition, oxalate deposition in bone may result in oxalate osteopathy, which in some cases may lead to accelerated bone maturation with reduced final height.⁶⁰

A multidisciplinary approach is needed to achieve the best possible management of primary hyperoxaluria. Patients and their families may find patient advocacy groups helpful

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on September 13, 2013. For personal use only. No other uses without permission.

(e.g., the Oxalosis and Hyperoxaluria Foundation [www.ohf.org], OxalEurope [www.oxaleurope.org], and Orphanet [www.orpha.net]).

TRANSPLANTATION

Since the liver is the sole organ responsible for glyoxylate detoxification, the excessive production of oxalate will continue as long as the native liver is present in patients with primary hyperoxaluria type 1. Thus, preemptive liver transplantation⁶¹ to avoid the complications of systemic oxalosis would appear to be a logical approach, with the surgery planned before the occurrence of stage 4 chronic kidney disease (estimated GFR, 15 to 30 ml per minute per 1.73 m²); this approach does raise ethical issues, given the risk of death associated with the procedure.62 Kidney transplantation without liver transplantation confers a very high risk of recurrence.6,35 Combined liver and kidney transplantation is therefore the treatment of choice for these patients.^{6,35,63,64} Kidney transplantation alone may be considered on an individual basis, such as in adults with confirmed responsiveness to pyridoxine. Dual transplantation is a reasonable choice for patients with stage 4 chronic kidney disease, since oxalate retention increases rapidly at this stage of renal dysfunction. In patients with stage 5 chronic kidney disease (estimated GFR, less than 15 ml per minute per 1.73 m²), sequential transplantation, starting with the liver, makes sense because the presence of a new, unaffected liver may permit the use of aggressive dialysis before renal transplantation, which may mobilize some of the systemic oxalate burden^{65,66} and protect the new kidney. Sequential transplantation is also an option when a suitable kidney is not available. Most studies have used transplants from deceased donors, but a living donor for split-liver and kidney transplantation may be considered.

The U.S. registry data on primary hyperoxaluria indicate a 5-year survival rate of 45% for kidney transplantation alone and 64% for dual kidney and liver transplantation³⁵; the corresponding rates for children are 14% and 76%,⁶ with the low rate of survival after kidney transplantation alone perhaps reflecting the severity of early-onset disease. Hemodialysis or hemofiltration is recommended during and after organ transplantation for recipients who have a heavy systemic oxalate burden, insufficient urine output, or both.³⁹ There is limited experience with organ transplantation in patients with primary hyperoxaluria type 2; the ubiquitous tissue distribution of GRHPR favors kidney transplantation, but some transplant recipients have undergone oxalate-related graft loss.⁴² Primary hyperoxaluria type 3 has not been associated with ESRD thus far.

Urinary oxalate excretion may remain elevated for many years after transplantation³⁵ because of the slow resolubilization of systemic calcium oxalate and may lead to nephrocalcinosis or renal calculi in the transplant. Consequently, the transplanted kidney must be protected through forced fluid intake and the use of crystallization inhibitors.

The specific multidisciplinary expertise and financial resources required for transplantation and subsequent management of primary hyperoxaluria are often not available in developing countries. As a result, many patients in these countries die, and for infants with severe primary hyperoxaluria, treatment may be withheld or withdrawn.⁶²

FUTURE THERAPEUTIC DEVELOPMENTS

Animal models have been developed for primary hyperoxaluria types $1,^{67}$ $2,^{68}$ and 3 (Salido E, Universidad La Laguna, Tenerife, Spain: personal communication). These models do not have the same phenotype as affected humans but are useful in the evaluation of treatments. The underlying problem in primary hyperoxaluria is not the enzyme deficiency itself but the accumulation of precursors, requiring replacement of liver tissue that is sufficient to overcome residual enzyme inactivity. Cell therapy, in which the liver is repopulated with normal hepatocytes, has been shown to be effective in Agxt knockout mice.69,70 However, there are still considerable difficulties in clinical applications of this approach to reduce the proliferation of host hepatocytes while boosting that of the transplanted cells. Hepatocyte transplantation has recently been suggested as a potential bridge to orthotopic liver transplantation in patients with primary hyperoxaluria type 1, but this procedure requires standard immunosuppressive therapy and does not fully correct the enzyme deficit.71 Gene transfer with the use of adeno-associated virus may be an attractive therapeutic option, but the problem of inducing ade-

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on September 13, 2013. For personal use only. No other uses without permission.

Table 2. Features and Treatment of the Inherited Primary Hyperoxalurias.				
Feature	Type 1	Туре 2	Туре 3	
Chromosomal location	2q37.3	9p13.2	10q24.2	
Age at onset	All ages, although mostly in childhood	All ages	All ages	
Presentation	Calcium oxalate renal stones, nephrocalcinosis, renal failure	Calcium oxalate renal stones	Calcium oxalate renal stones	
Treatment				
Supportive treatment	Hydration, citrate, pyridoxine	Hydration, citrate	Hydration, citrate	
Transplantation	Liver and kidney	Kidney	Not required — no reported cases of renal failure to date	

quate expression in addition to neutralizing antibodies must be overcome. Although the inhibition of glycolate oxidase could lead to substrate reduction, no suitable inhibitor has been identified as yet. Finally, the identification of pharmacologic chaperones to restore correct protein folding may be applicable to some genotypes. Recognition of such molecules depends on the use of high-throughput screens.^{10,72}

SUMMARY

Primary hyperoxaluria should be considered in any patient with a history of recurrent calcium oxalate stones, nephrocalcinosis, or both (Table 2).

REFERENCES

 Lepoutre C. Calculs multiples chez un enfant: Infiltration du parenchyme rénal par des dépôts cristallins. J Urol 1925; 20:424.

2. Mulay SR, Kulkarni OP, Rupanagudi KV, et al. Calcium oxalate crystals induce renal inflammation by NLRP3-mediated IL-1 β secretion. J Clin Invest 2013;123: 236-46.

3. Hoppe B, Kemper MJ, Bökenkamp A, Portale AA, Cohn RA, Langman CB. Plasma calcium oxalate supersaturation in children with primary hyperoxaluria and end-stage renal failure. Kidney Int 1999; 56:268-74.

4. Cochat P, Deloraine A, Rotily M, Olive F, Liponski I, Deries N. Epidemiology of primary hyperoxaluria type 1. Nephrol Dial Transplant 1995;10:Suppl 8:3-7.

5. van Woerden CS, Groothoff JW, Wanders RJ, Davin JC, Wijburg FA. Primary hyperoxaluria type 1 in the Netherlands: prevalence and outcome. Nephrol Dial Transplant 2003;18:273-9.

6. Harambat J, van Stralen KJ, Espinosa L, et al. Characteristics and outcomes of children with primary oxalosis requiring renal replacement therapy. Clin J Am Soc Nephrol 2012;7:458-65.

7. Kamoun A, Lakhoua R. End-stage renal disease of the Tunisian child: epidemiology, etiologies, and outcome. Pediatr Nephrol 1996;10:479-82.

8. Zhang X, Roe SM, Pearl LH, Danpure CJ. Crystallization and preliminary crystallographic analysis of human alanine: glyoxylate aminotransferase and its polymorphic variants. Acta Crystallogr D Biol Crystallogr 2001;57:1936-7.

9. Cregeen DP, Williams EL, Hulton SA, Rumsby G. Molecular analysis of the glyoxylate reductase (GRHPR) gene and description of mutations underlying primary hyperoxaluria type 2. Hum Mutat 2003; 22:497.

10. Behnam JT, Williams EL, Brink S, Rumsby G, Danpure CJ. Reconstruction of human hepatocyte glyoxylate metabolic pathways in stably transformed Chinesehamster ovary cells. Biochem J 2006; 394:409-16.

11. Baker PRS, Cramer SD, Kennedy M, Assimos DG, Holmes RP. Glycolate and glyoxylate metabolism in HepG2 cells. Am J Physiol Cell Physiol 2004;287:C1359-C1365.

12. Belostotsky R, Seboun E, Idelson GH, et al. Mutations in DHDPSL are responsi-

Once the diagnosis has been confirmed by genetic testing, aggressive supportive treatment is indicated, followed by an appropriate organtransplantation strategy if renal function is declining. Future therapeutic developments are aimed at correcting the underlying defects without exposing patients to the lifelong risks associated with organ transplantation.

Dr. Cochat reports receiving consulting fees from Lucane Pharma and Advicenne Pharma, providing expert testimony for Centre d'Investigation Clinique–Robert-Debré, and receiving support for travel expenses through his institution from Raptor Pharmaceutical, Novartis, Pfizer, Fresenius Medical Care, Amgen, Baxter, Merck Serono, Astellas, and Sandoz. No other potential conflict of interest relevant to this article was reported.

Disclosure forms provided by the authors are available with the full text of this article at NEJM.org.

ble for primary hyperoxaluria type III. Am J Hum Genet 2010;87:392-9.

13. Riedel TJ, Johnson LC, Knight J, Hantgan RR, Holmes RP, Lowther WT. Structural and biochemical studies of human 4-hydroxy-2-oxoglutarate aldolase: implications for hydroxyproline metabolism in primary hyperoxaluria. PLoS One 2011; 6(10):e26021.

14. Belostotsky R, Pitt JJ, Frishberg Y. Primary hyperoxaluria type III — a model for studying perturbations in glyoxylate metabolism. J Mol Med (Berl) 2012;90: 1497-504.

15. Riedel TJ, Knight J, Murray MS, Milliner DS, Holmes RP, Lowther WT. 4-Hydroxy-2-oxoglutarate aldolase inactivity in primary hyperoxaluria type 3 and glyoxylate reductase inhibition. Biochim Biophys Acta 2012;1822:1544-52.

16. Rumsby G. An overview of the role of genotyping in the diagnosis of the primary hyperoxalurias. Urol Res 2005;33: 318-20.

17. Fargue S, Lewin J, Rumsby G, Danpure CJ. Four of the most common mutations in primary hyperoxaluria type 1 unmask the cryptic mitochondrial targeting sequence of alanine:glyoxylate amino-

N ENGLJ MED 369;7 NEJM.ORG AUGUST 15, 2013

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on September 13, 2013. For personal use only. No other uses without permission.

transferase encoded by the polymorphic minor allele. J Biol Chem 2013;288:2475-84.

18. Rumsby G. Primary hyperoxaluria database (http://www.uclh.nhs.uk/phmd).
19. Williams EL, Acquaviva C, Amoroso A, et al. Primary hyperoxaluria type 1: update and additional mutation analysis of the AGXT gene. Hum Mutat 2009;30:910-7.

20. Benhaj Mbarek I, Abroug S, Omezzine A, et al. Selected AGXT gene mutations analysis provides a genetic diagnosis in 28% of Tunisian patients with primary hyperoxaluria. BMC Nephrol 2011;12:25.
21. Santana A, Salido E, Torres A, Shapiro LJ. Primary hyperoxaluria type 1 in the Canary Islands: a conformational disease due to I244T mutation in the P11L-containing alanine:glyoxylate aminotransferase. Proc Natl Acad Sci U S A 2003;100: 7277-82.

22. Monico CG, Rossetti S, Olson JB, Milliner DS. Pyridoxine effect in type I primary hyperoxaluria is associated with the most common mutant allele. Kidney Int 2005;67:1704-9.

23. Harambat J, Fargue S, Acquaviva C, et al. Genotype-phenotype correlation in primary hyperoxaluria type 1: the p.Gly170Arg AGXT mutation is associated with a better outcome. Kidney Int 2010; 77:443-9.

24. Fargue S, Rumsby G, Danpure CJ. Multiple mechanisms of action of pyridoxine in primary hyperoxaluria type 1. Biochim Biophys Acta 2013;1832:1776-83.
25. Frishberg Y, Rinat C, Shalata A, et al. Intra-familial clinical heterogeneity: absence of genotype-phenotype correlation in primary hyperoxaluria type 1 in Israel. Am J Nephrol 2005;25:269-75.

26. Williams EL, Bockenhauer D, van't Hoff WG, et al. The enzyme 4-hydroxy-2-oxoglutarate aldolase is deficient in primary hyperoxaluria type 3. Nephrol Dial Transplant 2012;27:3191-5.

27. Beck BB, Baasner A, Buescher A, et al. Novel findings in patients with primary hyperoxaluria type III and implications for advanced molecular testing strategies. Eur J Hum Genet 2013;21:162-72.

28. Monico CG, Rossetti S, Belostotsky R, et al. Primary hyperoxaluria type III gene HOGA1 (formerly DHDPSL) as a possible risk factor for idiopathic calcium oxalate urolithiasis. Clin J Am Soc Nephrol 2011; 6:2289-95.

29. Lieske JC, Monico CG, Holmes WS, et al. International registry for primary hyperoxaluria. Am J Nephrol 2005;25: 290-6.

30. van der Hoeven SM, van Woerden CS, Groothoff JW. Primary hyperoxaluria type 1, a too often missed diagnosis and potentially treatable cause of end-stage renal disease in adults: results of the Dutch cohort. Nephrol Dial Transplant 2012;27:3855-62.

31. Punjabi OS, Riaz K, Mets MB. Crystal-

line retinopathy in primary hyperoxaluria. J AAPOS 2011;15:214-6.

32. Bakshi NA, Al-Zahrani H. Bone marrow oxalosis. Blood 2012;120:8.

33. Mookadam F, Smith T, Jiamsripong P, et al. Cardiac abnormalities in primary hyperoxaluria. Circ J 2010;74:2403-9.

34. Beck BB, Hoyer-Kuhn H, Göbel H, Habbig S, Hoppe B. Hyperoxaluria and systemic oxalosis: an update on current therapy and future directions. Expert Opin Investig Drugs 2013;22:117-29.

35. Bergstralh EJ, Monico CG, Lieske JC, et al. Transplantation outcomes in primary hyperoxaluria. Am J Transplant 2010;10: 2493-501.

36. van Woerden CS, Groothoff JW, Wijburg FA, Annink C, Wanders RJA, Waterham HR. Clinical implications of mutation analysis in primary hyperoxaluria type 1. Kidney Int 2004;66:746-52.

37. Milliner DS, Wilson DM, Smith LH. Phenotypic expression of primary hyperoxaluria: comparative features of types I and II. Kidney Int 2001;59:31-6.

38. Milliner DS. The primary hyperoxalurias: an algorithm for diagnosis. Am J Nephrol 2005;25:154-60.

39. Cochat P, Hulton SA, Acquaviva C, et al. Primary hyperoxaluria Type 1: indications for screening and guidance for diagnosis and treatment. Nephrol Dial Transplant 2012;27:1729-36.

40. Daudon M, Estepa L, Lacour B, Jungers P. Unusual morphology of calcium oxalate calculi in primary hyperoxaluria. J Nephrol 1998;11:Suppl 1:51-5.

41. Daudon M, Jungers P, Bazin D. Peculiar morphology of stones in primary hyperoxaluria. N Engl J Med 2008;359:100-2.
42. Hoppe B. An update on primary hyperoxaluria. Nat Rev Nephrol 2012;8:467-75.
43. Barratt TM, Kasidas GP, Murdoch I, Rose GA. Urinary oxalate and glycolate excretion and plasma oxalate concentration. Arch Dis Child 1991;66:501-3.

44. Dietzen DJ, Wilhite TR, Kenagy DN, Milliner DS, Smith CH, Landt M. Extraction of glyceric and glycolic acids from urine with tetrahydrofuran: utility in detection of primary hyperoxaluria. Clin Chem 1997;43:1315-20.

45. Williams HE, Smith LH. The identification and determination of glyceric acid in human urine. J Lab Clin Med 1968;71: 495-500.

46. Rumsby G, Sharma A, Cregeen DP, Solomon LR. Primary hyperoxaluria type 2 without L-glycericaciduria: is the disease under-diagnosed? Nephrol Dial Transplant 2001;16:1697-9.

47. Rumsby G. Experience in prenatal diagnosis of primary hyperoxaluria type 1. J Nephrol 1998;11:Suppl 1:13-4.

48. Fargue S, Harambat J, Gagnadoux MF, et al. Effect of conservative treatment on the renal outcome of children with primary hyperoxaluria type 1. Kidney Int 2009;76:767-73.

49. Marangella M, Bagnis C, Bruno M, Vitale C, Petrarulo M, Ramello A. Crystallization inhibitors in the pathophysiology and treatment of nephrolithiasis. Urol Int 2004;72:Suppl 1:6-10.

50. Sikora P, von Unruh GE, Beck B, et al. [13C2]oxalate absorption in children with idiopathic calcium oxalate urolithiasis or primary hyperoxaluria. Kidney Int 2008;73: 1181-6.

51. Hatch M, Gjymishka A, Salido EC, Allison MJ, Freel RW. Enteric oxalate elimination is induced and oxalate is normalized in a mouse model of primary hyperoxaluria following intestinal colonization with Oxalobacter. Am J Physiol Gastrointest Liver Physiol 2011;300:G461-G469.

52. Hoppe B, Groothoff JW, Hulton SA, et al. Efficacy and safety of Oxalobacter formigenes to reduce urinary oxalate in primary hyperoxaluria. Nephrol Dial Transplant 2011;26:3609-15.

53. Pais VM Jr, Assimos DG. Pitfalls in the management of patients with primary hyperoxaluria: a urologist's perspective. Urol Res 2005;33:390-3.

54. Al-Abadi E, Hulton SA. Extracorporeal shock wave lithotripsy in the management of stones in children with oxalosis — still the first choice? Pediatr Nephrol 2013;28:1085-9.

55. Blomgren PM, Connors BA, Lingeman JE, Willis LR, Evan AP. Quantitation of shock wave lithotripsy-induced lesion in small and large pig kidneys. Anat Rec 1997;249:341-8.

56. Straub M, Gschwend J, Zorn C. Pediatric urolithiasis: the current surgical management. Pediatr Nephrol 2010;25: 1239-44.

57. Illies F, Bonzel KE, Wingen AM, Latta K, Hoyer PF. Clearance and removal of oxalate in children on intensified dialysis for primary hyperoxaluria type **1.** Kidney Int 2006;70:1642-8.

58. Hoppe B, Kemper MJ, Bökenkamp A, Langman CB. Plasma calcium-oxalate saturation in children with renal insufficiency and in children with primary hyperoxaluria. Kidney Int 1998;54:921-5.

59. Sahin G, Acikalin MF, Yalcin AU. Erythropoietin resistance as a result of oxalosis in bone marrow. Clin Nephrol 2005;63:402-4.

60. Bacchetta J, Fargue S, Boutroy S, et al. Bone metabolism in oxalosis: a singlecenter study using new imaging techniques and biomarkers. Pediatr Nephrol 2010;25:1081-9.

61. Kemper MJ, Nolkemper D, Rogiers X, et al. Preemptive liver transplantation in primary hyperoxaluria type 1: timing and preliminary results. J Nephrol 1998;11: Suppl 1:46-8.

62. Cochat P, Groothoff J. Primary hyperoxaluria type 1: practical and ethical issues. Pediatr Nephrol 2013 March 14 (Epub ahead of print).

N ENGLJ MED 369;7 NEJM.ORG AUGUST 15, 2013

657

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on September 13, 2013. For personal use only. No other uses without permission.

63. Jamieson NV. The results of combined liver/kidney transplantation for primary hyperoxaluria (PH1) 1984-1997: the European PH1 transplant registry report. J Nephrol 1998;11:Suppl 1:36-41.

64. Lorenzo V, Alvarez A, Torres A, Torregrosa V, Hernández D, Salido EC. Presentation and role of transplantation in adult patients with type 1 primary hyperoxaluria and the I244T AGXT mutation: single-center experience. Kidney Int 2006; 70:1115-9.

65. Malla I, Lysy PA, Godefroid N, et al. Two-step transplantation for primary hyperoxaluria: cadaveric liver followed by living donor related kidney transplantation. Pediatr Transplant 2009;13:782-4.
66. Mor E, Nesher E, Ben-Ari Z, et al. Sequential liver and kidney transplantation from a single living donor in two young adults with primary hyperoxaluria type 1. Liver Transpl 2013;19:646-8.

67. Salido EC, Li XM, Lu Y, et al. Alanineglyoxylate aminotransferase-deficient mice, a model for primary hyperoxaluria that responds to adenoviral gene transfer. Proc Natl Acad Sci U S A 2006;103: 18249-54.

68. Knight J, Holmes RP, Cramer SD, Takayama T, Salido E. Hydroxyproline metabolism in mouse models of primary hyperoxaluria. Am J Physiol Renal Physiol 2012;302:F688-F693.

69. Jiang J, Salido EC, Guha C, et al. Correction of hyperoxaluria by liver repopulation with hepatocytes in a mouse model

of primary hyperoxaluria type-1. Transplantation 2008;85:1253-60.

70. Guha C, Yamanouchi K, Jiang J, et al. Feasibility of hepatocyte transplantationbased therapies for primary hyperoxalurias. Am J Nephrol 2005;25:161-70.

 Beck BB, Habbig S, Dittrich K, et al. Liver cell transplantation in severe infantile oxalosis — a potential bridging procedure to orthotopic liver transplantation? Nephrol Dial Transplant 2012;27:2984-9.
 Hopper ED, Pittman AMC, Fitzgerald MC, Tucker CL. In vivo and in vitro examination of stability of primary hyperoxaluria-associated human alanine:glyoxylate aminotransferase. J Biol Chem 2008;283: 30493-502.

Copyright © 2013 Massachusetts Medical Society.

The New England Journal of Medicine

Downloaded from nejm.org by LUIGI GRECO on September 13, 2013. For personal use only. No other uses without permission.